DITERPENOIDS FROM VIGUIERA INSIGNIS

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Abstract—The isolation of several known diterpenoids and one new diterpene diol from aerial parts of Viguiera insignis is reported.

INTRODUCTION

Several Viguiera species have yielded sesquiterpene lactones [1-6], diterpenoids [7] or both [8]. In continuation of our chemical investigation on this and related genera, we now report the isolation and characterization of the diterpenes from Viguera insignis Miranda.

RESULTS AND DISCUSSION

The less polar fractions of the chromatography of the chloroform extract of V. insignis contained several acidic compounds. This mixture of acids was methylated and separated into its components by Si gel chromatography. The components, in order of increasing polarity, were: ent-kaur-9(11)-16-dien-19-oic acid (1) [9], stach-15-en-19-oic acid (3) [8] and ent-kaur-16-en-19-oic acid (2) [10]. The identification of these compounds was carried out by physical constants, spectroscopic methods and direct comparison with authentic samples. The diterpene alcohol, stach-15-en-3 α -ol (4) was isolated from these fractions and was identified by its spectroscopic characteristics [11].

The more polar fractions afforded two isomeric diols derived from stach-15-ene. One of these displayed physical and spectroscopic data identical with those reported for (+)-stach-15-en-3 α ,19-diol (5). This was converted into the known keto-aldehyde (6) and the ethylidene derivative (7) [12], thus confirming its structure. The second diol (8) C₂₀H₃₂O₂ (elemental analysis and mass spectrometry) also had features characteristic of a stachene derivative: a cis disubstituted double bond shown in ¹H NMR as an AB system (δ 5.70, 5.53; J = 5.7 Hz) which shifted upfield on epoxidation in 9 and 10, and four quaternary methyl groups (see Table 1). The signals at δ 0.98 and 0.79 are in good agreement with a 4,4'-gemdimethyl group adjacent to a 3-equatorial hydroxyl group [13] and the paramagnetic shift of Me-10 in the dihydro derivatives on hydrogenation in 11 and 12 indicates that the C-20 methyl group is present as such [14]. The presence of the second hydroxyl function on ring C was confirmed by the transformation of 8 to the derivatives

13–15. The absence of a large coupling constant of the signal in ¹H NMR of the second carbinolic proton $(W_{1/2} = 7 \text{ Hz})$ suggests an axial orientation of the hydroxyl group. The experiments with Eu(fod)₃ d demonstrated that the C-17 methyl showed a remarkable downfield shift concordant only with a 12- β -ol (see Table 1). The structure and stereochemistry proposed for this diol was confirmed by X-ray diffraction and the stereostructure is depicted in Fig. 1. A complete list of refined coordinates is deposited at the Cambridge Crystallographic Data Centre.

R³ R⁴ R⁵ R R2 н COOH н н ОН Me н 5 OH CH2OH =0 CH= O 6 н ОН ОН 0 H В – ероху 10 OAc OAC B - epoxy 1.1 ОН ОН dihydro 12 OAC Me OAC 15, 16 dihydro 13 OAC Me ОН н OAc 14 Me OAC н 15 =0 = 0 Me

^{*}Contribution No. 613.

Table 1 ¹H NMR data of compounds 8-15 (80 MHz, CDCl₃)*

| | | | | lable 1 'H'N | MK data of c | lable 1 'H NMK data of compounds 8-15 (80 MHz, | | CDCl ₃)* | | | |
|------|------------------------------|---------------------------|-------------------------|--------------------------|-------------------------|--|-------------------------|-----------------------|----------------------|------------------------|-----------------|
| | ∞ | 8a↑ | 8 ₽ ‡ | \$c† | 6 | 10 | 11 | 12 | 13 | 14 | 15 |
| 1-3 | 3 18 dd (10, 54) | | 4 90 dd (10, 5 4) | 5 72 dd (10, 5 4) | 3 18 dd (10, 5 4) | 4 46 dd (10, 5 4) | 3 16 dd (10, 5 4) | 4 45 dd (10, 5 4) | 4 48 dd (10, 5 4) | 4 48 dd (10, 5 4) | |
| I-12 | H-12 $365 m$ $(W_{1,2} = 7)$ | $4\ 30\ m (W_{1/2} = 7)$ | $500 m$ $(W_{1/2} = 7)$ | $5.68 m$ $(W_{1/2} = 7)$ | $3.76 m (W_{1/2} = 7)$ | $495 m $ $(W_{1/2} = 7)$ | $3.48 m (W_{1/2} = 7)$ | $467 m (W_{1/2} = 7)$ | | $483 m (W_{1/2} = 7)$ | 1 |
| I-15 | 571 d (57) | | 6 10 d (5 7) | 6 25 d (5 7) | 3 34 <i>d</i> (3 0) | 3 37 d | 1 | ŀ | | 5 79 d (5 7) | 6 10 d (5 7) |
| I-16 | 5 53 d (5 7) | | 5 90 d (5 7) | 6 05 d (5.7) | 2 <i>97 d</i> (3 0) | 3 04 d (3 0) | I | I | 5 54 <i>d</i> (5 7) | 5 55 d (5 7) | 5 64 d (5 7) |
| 1-17 | 1 05 5 | | 17.5 | 2 10 s | 1 10 s | 1 00 s | 1.02 s | 093 s | | 0.95 \$ | 1 11 s |
| -10 | 0 79 s 0 79 s | | 1 /0 s 1 65 s | 2 10 s 2 04 s | 1 00 s 0 81 s | 0 88 s 0 88 s | 2 66 0 0 79 s | 0.86 s 0.86 s | | 086 s 086 s | 111 5 |
| 1-20 | s 69 0 | | 118, | 1 40 5 | 091 s | 0.93 \$ | 0918 | 093 s | | 0.71.5 | 095 8 |
| | | | | | | | | | | | |

*Coupling constants (Hz) in parentheses †Addition of (8a) 0 105, (8b) 0 222 and (8c) 0 327 mol Eu (fod)₃ per mol substrate

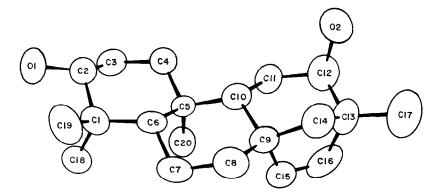


Fig 1

The results obtained are in agreement with the similarities observed in the terpenoids of some sections of Viguiera and Helianthus [15], which are placed in the same subtribe (Helianthineae) The diterpenoid acids are widespread in these genera

EXPERIMENTAL

Mps are uncorr

Aerial parts (3 1 kg) of V insigns, (collected on 19 November 1979, ca 16 km SSE of Izucar de Matamoros, Puebla, Hwy 190, voucher AOH 0022, MEXU 320489) were extracted with CHCl₃ affording 133 g residue. This was fractionated over tonsil eluting with hexane–CHCl₃ (9 1) and with CHCl₃–EtOAc (8.2) yielding 72 3 and 42 1 g residue, respectively. Part of the non-polar residue (20 g) was chromatographed on Si gel using hexane–EtOAc (9 1). The first fractions were combined and evaporated, leaving a residue (1 3 g) which was treated with CH₂N₂. The mixture containing the methyl esters was chromatographed on Si gel. The fractions eluted with hexane–C₆H₆ (20 1) afforded 1–3 as methyl esters. From the fractions eluted with hexane–EtOAc (9 2), was isolated 4

The polar residue was chromatographed on Si gel and elution with CHCl₃–Me₂CO (20 1) gave (+)-stach-15-en-3 α ,19-diol (5) (1 21 g) Subsequent fractions eluted with CHCl₃–Me₂CO (4 1) gave 1 73 g (+)-stach-15-en-3 α ,12 β -diol (8) Mp 204–205° [α] $_{\rm D}^{\rm 25}$ + 8 73° (MeOH, c 0.126) IR $\nu_{\rm max}^{\rm nujol}$ cm $^{-1}$ 3360, 1460, 1412, MS m/z (rel int) 304 [M] $^+$ (3 8), 286 (2 3) 110 (24 6), 119 (34 6), 107 (100), 105 (65 3), 93 (69 2), 91 (74 6), 81 (55 4), 43 (61 5). (Found. C, 78 71, H, 10 60, Q, 10 53 C₂₀H₃₂Q₂ requires C, 78 89, H, 10 59, O, 10 51 %) This substance was submitted to X-ray analysis, giving the following general parameters a (A) = 12 221 (2), b(A) = 15 166 (2), c(A) = 19 471 (5) Space group P2₁2₁2₁ ρ _{calc} (mg/m³) = 1 120 (1), wavelength, MoK α = 0 71073 A

Derivatives of 5 150 mg 5 in Me₂CO was treated with Jones reagent at 0° for 10 min. Usual work-up gave 6 (130 mg), mp $108-109^{\circ}$, lit [12] $109-110^{\circ}$ 180 mg 5 in Et₂O was shaken with paraldehyde (1 2 ml) and HCl (0 4 ml) to give, after usual work-up, 140 mg 7, mp $120-121^{\circ}$, lit [12] $120-121^{\circ}$

Derivatives of **8** Acetylation 213 mg **8** was acetylated to give, after TLC (eluted with hexane–EtOAc, 4 1) the diacetate (14) (89 1 mg), mp $125-127^{\circ}$, IR $v_{\rm max}^{\rm CHCl_3}$ cm $^{-1}$ 1731, 1370, 1250, MS m/z (rel int) 388 [M] $^+$ (47), 328 (25 1), 268 (4 2), 253 (6.2), 145 (23.9), 133 (92 3), 119 (44 1), 107 (38 2), 106 (50 9), 91 (57 7), 43 (100) (Found C, 73 95, H, 9 48, O, 16 70 C₂₄H₃₆O₄ requires C, 74 19, H, 9 34, O, 16 47%) and the monoacetate (13) (57 5 mg), mp $137-139^{\circ}$ IR $v_{\rm max}^{\rm CHCl_3}$ cm $^{-1}$ 3600, 1730, 1350, 1250 MS m/z (rel int) 346 [M] $^+$ (1 0), 330 (5 1), 316 (0 5), 270 (20 1), 255 (12 5), 107 (25 0), 105 (26 4), 93 (34 6), 81 (25 0), 43 (100)

Epoxidation Treatment of **8** (110 mg) in CH₂Cl₂ with mcpba (100 mg) afforded 95 mg 9, mp 194–196°, IR $\nu_{\rm mc}^{\rm CHCl_3}$ cm $^{-1}$ 3420, 1450, 1030, MS m/z (rel int) 320 [M] $^+$ (20 1), 302 (4 7), 287 (5 8), 250 (10 5), 249 (56 9), 161 (21 0), 159 (20 1), 147 (27.5), 145 (28 5), 135 (100), 123 (46 5), 107 (67 0), 105 (77 9), 93 (68 4), 91 (94 7), 79 (84 1)

Hydrogenation 139 mg **8** in MeOH was hydrogenated using Pd–C to afford 126 5 mg **11**, mp 198–199° $[\alpha]_{2}^{D5}$ – 24 53° (MeOH, c 0 163) IR $v_{\rm max}^{\rm CHCl_3}$ cm $^{-1}$ 3600, 1448, 1380, 1362 MS m/z (rel. int.) 306 [M] $^+$ (13 9), 288 (21 7), 273 (20 0), 231 (18 0), 136 (73 2), 121 (82 9), 109 (54 0), 197 (100), 105 (65 1), 79 (78 9), 43 (40.0) (Found C, 78 67; H, 11 23, O, 10 20 C₂₀H₃₄O₂ requires C, 78 38, H, 11 18, O, 10 44%)

Oxidation Jones oxidation of 121 1 mg **8** gave 74 3 mg **15** after TLC (eluted with CHCl₃–Me₂CO, 9 1), mp 92–94°, IR $v_{\rm CHCl_3}^{\rm CHCl_3}$ cm $^{-1}$ 1770, 1440, 1380, 1360 MS m/z (rel int) 300 [M] $^+$ (58 3), 272 (8.1), 148 (12 1), 145 (11 5), 119 (32 2), 107 (38 2), 106 (68 8), 105 (53 3), 93 (100), 91 (84 1), 79 (51 1), 77 (58 0), 67 (38 2)

Conversion of 14 to 10 90 mg 14 in CH₂Cl₂ was oxidized with mcpba to give 70 mg 10, mp 178–180°, IR $v_{\rm mc}^{\rm CHCl_3}$ cm $^{-1}$ 1740, 1460, 1372, 1250 MS m/z (rel int). 404 [M] $^+$ (2 6), 344 (4 8), 269 (5 5), 255 (6 0), 161 (7 6), 147 (10 0), 135 (26 4), 131 (10 9), 121 (19 3), 107 (15.7), 105 (20 1), 95 (23 7), 43 (100)

Conversion of 14 to 12. Catalytic hydrogenation of 14 (95 1 mg) in EtOAc afforded 12 (87 3 mg), 135–136° IR $v_{\rm max}^{\rm CHCl_3}$ cm $^{-1}$ 1720, 1450, 1365, 1250 MS m/z (rel int.): 390 [M] $^+$ (0.5), 330 (9 3), 316 (1 8) 302 (1 0), 288 (1 0), 270 (43 4), 255 (47 9), 226 (20 0), 136 (30 1), 121 (39 8), 107 (40 0), 93 (35.2), 91 (35 0), 43 (100). (Found C, 73 56; H, 9 75, O, 16 30 C₂₄H₃₈O₄ requires C, 73 80, H, 9 81, O, 16 39 %)

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REFERENCES

- 1 Romo de Vivar, A, Guerrero, C, Díaz, E, Bratoeff, E. and Jiménez, L (1976) Phytochemistry 15, 525
- Romo de Vivar, A., Delgado, G., Guerrero, C., Reséndiz, J. and Ortega, A. (1978) Rev. Latinoam Quim 9, 171
- 3 Guerrero, C, Santana, M and Romo, J, (1976) Rev Latinoam. Quim 7, 41
- 4 Ortega, A, Lara, R, Martínez, R and Díaz, E (1980) Phytochemistry 19, 1545

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5 Romo de Vivar, A, Bratoeff, E, Ontiveros, E, Lankin, D C and Bhacca, N S. (1980) Phytochemistry 19, 1795

- 6 Delgado, G, Romo de Vivar, A and Herz, W (1982) Phytochemistry 21, 1305
- 7 Bohlmann, F, Zdero, Ch and Mahanta, P (1977) Physochemistry 16, 1073
- 8 Bohlmann, F, Jakupovic, J, Ahmed, M, Grenz, M, Suding, H, Robinson, H and King, R M (1981) Phytochemistry 20, 113
- 9 Brieskorn, C H and Pohlman, E (1968) Tetrahedron Letters 5661
- 10 Henrick, C. A. and Jefferies, P. R. (1964). Aust. J. Chem. 17, 915
- 11 Baarschers, W H, Horn, D H S. and Johnson, L R F. (1962) J Chem Soc 4046
- 12 Lloyd, H A and Fales, H M (1967) Tetrahedron Letters 4891
- 13 Hanson, J R (1970) Tetrahedron 26, 2711
- 14 von Carstenn-Lichterfelde, C, Valverde, S and Rodríguez B (1974) Aust J Chem 27, 517
- 15 Ohno, N, Gersenzon, J, Neuman, P and Mabry, T J (1981) Phytochemistry 20, 2393